# FROZEN STATE TRANSITIONS IN RELATION TO FREEZE DRYING

# Y. H. Roos

Department of Food Technology, PO Box 27 (Viikki B), FIN-00014 University of Helsinki, Helsinki, Finland

# Abstract

Freeze-drying is used as a gentle dehydration method for heat sensitive materials especially in food and pharmaceutical industries. Most materials, including dissolved sugars in water, do not crystallise during freezing prior to freeze-drying. Supersaturated, freeze-concentrated solutions are amorphous materials and they solidify into a glassy state when their temperature is depressed to below the glass transition temperature,  $T_{\rm g}$ . Differential scanning calorimetry has been used to show that maximally freeze-concentrated sugar solutions, when properly frozen, show during heating a glass transition,  $T'_g$ , which is followed by ice melting endotherm with onset at  $T'_{\rm m}$ . Low molecular weight materials are difficult to freeze-dry as they have low  $T'_{\rm g}$ , and  $T'_{\rm m}$ slightly above  $T'_{z}$ . High molecular weight materials, such as carbohydrate polymers, exhibit improved dehydration characteristics and they have  $T'_g$  and  $T'_m$  at about the same temperature close to the melting point of pure water. The amorphous, glassy structure typical of freeze-dried materials is formed during prefreezing and retained after removal of ice and the unfrozen water from the freeze-concentrated material. Dehydration temperatures below  $T'_{e}$  allow removal of ice within the solid, glassy solutes, but temperatures above  $T'_m$  result in collapse. The frozen state transitions and properties of freeze-dried materials can be shown in state diagrams which are used to derive proper freeze-drying conditions and storage requirements for various materials.

Keywords: collapse, freeze-drying, glass transition, ice melting, water plasticisation

# Introduction

Freeze-drying is an important drying method providing a gentle dehydration of heat sensitive materials especially in food and pharmaceutical industries. A successful freeze-drying process retains the volume of the material which, in the dry state, is usually highly porous, brittle, and hygroscopic, but has excellent rehydration properties. However, the structure of a number of materials, in particular, that of those containing sugars may collapse during dehydration if the temperature of ice within the material is higher than the collapse temperature,  $T_c$ , of the material. The collapsed materials exhibit poor dehydration properties and loss of desired quality characteristics [1].

It is well known that materials which form eutectic solutions with water, such as salts, collapse if the freeze-drying temperature is higher than the eutectic melting temperature,  $T_e$ , e.g.,  $-22^{\circ}$ C for NaCl [2]. Such collapse results from the presence

of liquid water that is removed by evaporation. Obviously, eutectic materials exist as crystalline solids after freeze-drying. However, most organic and biological materials, including dissolved sugars in water, do not crystallise during freezing prior to freeze-drying. During the prefreezing step water is removed from solution as ice, and the solute phase becomes freeze-concentrated. Freeze-concentration increases with decreasing temperature and at temperatures below  $T_e$  the viscosity of the freeze-concentrated, unfrozen solute phase also increases, as, in addition to the lowering temperature, water is removed from the supersaturated solution as ice. At low temperatures the high viscosity also delays ice formation and the glassy state is formed before all water is removed from the solution as ice [3]. Ice formation ceases due to glass formation and, therefore, frozen materials at low temperatures often contain maximally freeze-concentrated, amorphous solutes [4, 5]. Differential scanning calorimetry (DSC) has been used to show that sugar solutions, when properly frozen, show during heating a glass transition which is followed by the ice melting endotherm [5]. The glass transition is observed at a material specific, initial concentration independent temperature which is usually referred to as the glass transition temperature of the maximally freeze-concentrated solute matrix,  $T'_{g}$ [4, 5]. The  $T'_g$  is followed by ice melting which has its onset at  $T'_m$  [5]. Low molecular weight materials with low  $T'_g$  have  $T'_m$  slightly above  $T'_g$  while high molecular weight materials, such as carbohydrate polymers, have  $T'_g$  and  $T'_m$  at about the same temperature close to the melting point of pure water [6]. Most maximally freezeconcentrated carbohydrate matrices contain about 80% w/w solids and 20% w/w unfrozen water and have  $T'_g$  well above the  $T_g$  of pure water of  $-135^{\circ}C$  [7, 8].

The frozen state transitions of various materials have proved to affect various material properties in cryopreservation and freeze-drying. The main emphasis of the present article will be on establishing relationships between the observed transitions of freeze-concentrated solids and water, and on the effects of the transitions on freeze-drying behaviour and physicochemical properties of the freeze-dried materials. Our premise is that the control of the prefreezing and freeze-drying processes, on the basis of knowledge of frozen state transitions, can be used to decrease quality changes and predict the drying behaviour of various materials.

# Phase and state behaviour of solutes and water

# Freezing of pure water

The principle of freeze-drying is based on the phase behaviour of pure water. It is well known that at atmospheric pressure the physical state of water is dependent on temperature, and water may exist as ice, liquid water, and water vapour. A reduction in pressure results in a decrease in boiling temperature until the triple point pressure of 6.104 mbar, at which all three phases are at equilibrium, is reached. The triple point temperature is 0.0099°C below which water may exist only as ice or water vapour. The main requirement for freeze-drying, i.e., removal of water by sublimation directly from the solid to the vapour state is that the temperature is below that of the triple point. This can be achieved by lowering temperature or pressure. In common freeze-drying methods the pressure is decreased to well below that of the triple point, and used to control temperature of the sublimating ice.

### Effects of solutes on ice formation

Solutes in water reduce vapour pressure, which in dilute solutions occurs according to the Raoult's law. The Raoult's law states that the vapour pressure of water in a solution, p, is equal to the product of vapour pressure of pure water,  $p_0$ , at the same temperature multiplied by its mole fraction,  $x_w$ , i.e.,  $p = p_0 x_w$ . The requirement for ice formation is that the vapour pressure of water is higher than the vapour pressure of ice,  $p_i$ , at the same temperature. The equilibrium melting temperature is the temperature where  $p = p_i$ . Therefore, the equilibrium melting temperature of water in solution is lower than that of pure ice, as the vapour pressure of water in solution equals that of ice at a lower temperature. However, ice formation in solutions results in freeze-concentration of solutes and there is a dynamic decrease in ice melting temperature with increasing ice formation due to the decreasing vapour pressure of unfrozen water. The various effects of solutes on ice formation and melting temperature are shown schematically in Fig. 1. It is also important to notice that ice formation above the eutectic temperature can be considered as equilibrium ice formation while nonequilibrium ice formation occurs at temperatures below  $T'_m$  where ice formation becomes kinetically restricted [9].

#### Glass transition

Supercooled liquids may exist as solid, glassy materials or in a viscous liquid state. The transition between the glassy and supercooled liquid states occurs over a glass transition temperature range. The transition exhibits properties of a secondorder phase transition, and it can be detected from changes in a number of thermal, mechanical, and dielectric properties [9]. The transition is often determined by DSC which detects a change in heat capacity,  $\Delta C_p$ , that occurs over the glass transition temperature range. In water-solute systems the transition occurs over a temperature range located between the glass transition temperatures of the pure components. The glass transition temperature is often given as the onset or midpoint temperature of the calorimetric glass transition temperature range. The glass transition temperature of pure water in various studies has been found to occur between -144 and  $-134^{\circ}C$  [9], but the values found for  $\Delta C_p$  over the transition have varied from 0.1 [10] to 1.94 J (g °C)<sup>-1</sup> [11]. However, the values often used as  $T_g$  and  $\Delta C_p$  for water have been -135°C [8] and 1.94 J (g °C)<sup>-1</sup> [11], respectively. Most freezedried materials consists of amorphous solids in which water acts as a plasticizer. The  $T_g$  of stable, low-moisture, freeze-dried materials must be well above storage temperature, as the materials cannot support their own weight and collapse due to liquid flow when stored at temperatures above  $T_g$  [9, 12]. The plasticizing effect of water is observed from a dramatic decrease of  $\tilde{T}_g$  with increasing water content of the material. Depression of  $T_g$  to below storage temperature, as a result of water plasticization, may cause stickiness, caking, collapse, crystallization of amorphous sugars, loss of encapsulated compounds, and other quality defects [9, 12].

#### State diagrams

State diagrams are used to characterize the physical state of amorphous, biological materials as a function of temperature and water content [13]. They are used to show the  $T_g$  as a function of water content and they often contain information of solubility limits and effects of freeze-concentration on transition temperatures.

State diagrams can be based on experimental measurements of  $T_g$ , solubility, and equilibrium melting temperature at various water contents [5–7, 9]. Completely amorphous materials with low water contents can be obtained, e.g., by freeze-drying or rapid cooling from melt. The water content of the material can be



Fig. 1 Ice formation in solutions. A. Ice formation in an eutectic solution occurs at equilibrium according to the melting temperature,  $T_m$ , curve until both water and solute crystallize at the eutectic temperature,  $T_e$ . B. Ice formation in biological materials occurs often according to the  $T_m$  curve at equilibrium, but as the solutes do not crystallize maximum ice formation occurs between the onset temperature of ice melting within the maximally freeze-concentrated solute phase,  $T'_m$ , and glass transition temperature of the maximally freeze-concentrated solute phase,  $T'_g$ . The  $T_g$  curve indicates the glass transition temperature at various solute concentrations, and the solute concentration for the maximally freeze-concentrated solute matrix is given by  $C'_g$ . Nonequilibrium ice formation, controlled by the viscosity of the freeze-concentrated unfrozen matrix and  $T_g$ , occurs in less than maximally freeze-concentrated solutions at temperatures below  $T'_g$ .

adjusted by rehumidification or addition of water. However, the transition temperatures are extremely sensitive to water and a homogeneous distribution of an exactly known water content is required. Determination of the frozen state transitions requires water contents higher than 20% w/w which is often the unfrozen water content of maximally freeze-concentrated solutes [9]. The glass transition curve can be predicted using the Gordon-Taylor [14] or Couchman-Karasz [15] equations which can be fitted to experimental  $T_g$  data. The equations define the  $T_g$  of the mixture of solids and water in terms of component  $T_g$  ( $T_{g1}$  and  $T_{g2}$  for solids and water, respectively), weight fractions ( $w_1$  and  $w_2$  for solids and water, respectively), and a constant, k, in the Gordon-Taylor Eq. (1) or the change in heat capacities,  $\Delta C_{p1}$  and  $\Delta C_{p2}$ , over the glass transitions of solids and water, respectively, in the Couchman-Karasz Eq. (2). The use of the Gordon-Taylor equation is often the most convenient, as the determination of  $\Delta C_{p1}$  can be difficult and the existing differences between the reported  $\Delta C_{p2}$  values.

$$T_{g} = \frac{w_{1}T_{g1} + kw_{2}T_{g2}}{w_{1} + kw_{2}} \tag{1}$$

$$\ln T_{g} = \frac{w_{1} \Delta C_{p1} \ln T_{g1} + w_{2} \Delta C_{p2} \ln T_{g2}}{w_{1} \Delta C_{p1} + w_{2} \Delta C_{p2}}$$
(2)

State diagrams have been reported for various sugars [7], polysaccharides [16], cereal proteins [17], and sugar-protein mixtures [18]. A typical example of state diagrams is that of sucrose (Fig. 2) showing the glass transition curve,  $T'_g$ ,  $T'_m$ , and equilibrium melting temperature,  $T_m$ . The solute concentration in a maximally freeze-concentrated matrix is given by  $C'_g$ .  $T_g$ ,  $T'_g$ ,  $T'_m$ ,  $C'_g$ , and constant k of the Gordon-Taylor equation are given for various sugars in Table 1.



Fig. 2 State diagram for sucrose showing experimental and predicted data for equilibrium ice melting temperature,  $T_m$ , onset temperature for ice melting within the maximally freeze-concentrated solute matrix,  $T'_m$ , glass transition temperature of the maximally freeze-concentrated solute matrix,  $T'_g$ , with concentration,  $C'_g$ , and glass transition temperature,  $T_o$ , as a function of solute concentration. After Roos [9]

Table 1 Glass transition temperatures,  $T_g$ , for various sugars and sugar alcohols. Maximum freeze-concentration occurs between  $T'_g$  and  $T'_m$  which are the glass transition temperature of the maximally freeze-concentrated solutes with solute concentration of  $C'_g$  and onset of ice melting within the maximally freeze-concentrated solute matrix, respectively. The constant, k, is that used in the Gordon-Taylor equation to obtain  $C'_g$  [7, 9].

Compound	$T_{a}/^{o}C^{a}$	$T_{\rm c}^{\prime \prime 0} C^{\rm a}$	T''/°C	k	C'/(% w/w)
Pentoses	<u> </u>				
Arabinose	-2	-66	-53	3.55	79.3
Ribose	-20	-67	-53	3.02	81.4
Xvlose	6	-65	-53	3.78	78.9
Hexoses					
Fructose	5	-57	-46	3.76	82.5
Fucose	26	62	-48	4.37	78,4
Galactose	30	-56	-45	4.49	80.5
Glucose	31	-57	-46	4.52	80.0
Mannose	25	58	-45	4.34	80.1
Rhamnose	-7	-60	-47	3.40	82.8
Sorbose	19	-57	-44	4.17	81.0
Disaccharides					
Lactose	101	-41	-30	6.56	81.3
Lactulose	79	-42	-32	5.92	82.0
Maltose	87	-42	-32	6.15	81.6
Melibiose	85	-42	-32	6.10	81.7
Sucrose	62	46	34	5.42	81.7
Trehalose	100	-40	30	6.54	81.6
Oligosaccharides					
Raffinose	70	-36	-28	5.66	84.1
Sugar alcohols					
Maltitol	39	-47	37	4.75	82.9
Sorbitol	9	-63	-49	3.35	81.7
Xylitol	-29	-72	57	2.76	80.2

<sup>a</sup>Onset of the glass transition temperature range

# **Structure formation**

# Ice formation

At freezing temperatures ice formation occurs according to the equilibrium melting temperature curve until the freezing temperature is sufficiently low to allow maximum ice formation. As shown by the state diagram for sucrose (Fig. 2) maximum ice formation occurs over the temperature range of  $T'_g$  to  $T'_m$ . Separation of

water as ice and the increase in solute concentration in the unfrozen phase results in an increase in viscosity. At temperatures below  $T'_m$  but above  $T'_g$  the viscosity of the unfrozen phase increases significantly and ice formation becomes extremely slow, as the viscosity increases to above  $10^7$  Pa s [5]. At temperatures below  $T'_g$  the viscosity of the unfrozen phase reaches the typical value of  $10^{12}$  Pa s for glassy materials and ice formation ceases.

Time-dependent ice formation is often observed in DSC scans of rapidly cooled solutions. Heating scans of rapidly cooled sugar solutions exhibit a glass transition at a relatively low temperature and a devitrification exotherm at a higher temperature which is followed by the ice melting endotherm. As shown schematically in Fig. 3, the exotherm occurs due to ice formation during reheating. No exotherm appears after proper annealing and maximum ice formation at  $T'_g < T < T'_m$ . Therefore, heating curves of annealed and maximally freeze-concentrated solutions show glass transition of the maximally freeze-concentrated solute phase and onset of ice melting at initial concentration-independent  $T'_g$  and  $T'_m$ , respectively.  $T'_g$  and  $T'_m$  values for various sugars are given in Table 1. It should be noticed that monosaccharides have low  $T'_m$  values and they are extremely difficult to freeze-dry. The  $T'_g$  and  $T'_m$  increase with increasing molecular weight and those of polysaccharides occur at about the same temperature slightly below the melting temperature of pure ice [6].

In freeze-drying the amorphous structure of freeze-concentrated solids formed in prefreezing must be retained during removal of ice and unfrozen water. The material temperature during sublimation is critical for a successful freeze-drying process. Temperatures below  $T'_g$  allow removal of ice within the solid, glassy solutes and there are practically no changes in volume. At temperatures above  $T'_m$  the unfrozen water content increases which results in water plasticization of the freezeconcentrated solutes. Therefore, the freeze-concentrated solute matrix is transformed into the liquid state. The plasticized solute matrix may flow above  $T'_m$  and



Fig. 3 Schematic representation of thermograms typical of rapidly cooled, nonannealed sugar solutions and annealed sugar solutions with maximum ice formation after proper annealing at  $T'_g < T < T'_m$ 

most materials are likely to foam and collapse in freeze-drying when dehydration temperatures higher than the respective  $T'_m$  are used. The state diagrams which show the  $T_g$  and frozen state transition temperatures as a function of concentration are extremely useful in evaluation of proper freeze-drying conditions.

#### Importance of vitrification

Maximally freeze-concentrated materials can be considered as solids which contain the unfrozen solute phase in the high viscosity solid state within the glass transition temperature range or in the glassy state and a separate ice phase. In freezedrying the unfrozen solids can support their own weight when the sublimation temperature of ice is kept below  $T'_m$  and no foaming or collapse are observed (Fig. 4). After removal of ice and unfrozen water the glassy state of solids in the dehydrated part of the material can be maintained when the  $T_g$  of the material is higher than material temperature. The same requirement applies after dehydration, as collapse of the dehydrated solids occurs if the material is stored above its  $T_g$  (Fig. 4). The dehydrated materials may also collapse due to water sorption from the surrounding atmosphere, which results in water plasticization and a possible decrease of the  $T_g$ to below ambient temperature. Several monosaccharides cannot be freeze-dried because their  $T'_m$  values are lower than practically achievable sublimation temperatures or low anhydrous  $T_g$  (Table 1). The low material  $T_g$  and high hygroscopicity are the main causes of collapse during dehydration or collapse occurring after suc-



Fig. 4 Effects of vitrification on the structure of freeze-dried materials. Sublimation temperatures lower than the onset temperature of ice melting after maximum ice formation,  $T'_m$ , allow removal of ice without collapse. Frozen layer temperatures higher than  $T'_m$  during freeze-drying cause plasticization of freeze-concentrated solutes which cannot support their own weight and collapse. Successfully freeze-dried materials may collapse when stored at temperatures higher than the glass transition temperature,  $T_g$ , of the material or as a result of water sorption and plasticization which may depress the  $T_g$  to below ambient temperature

543

cessful freeze-drying. However, it should be noticed that collapse processes are time-dependent with relaxation times decreasing with increasing temperature above  $T_g$  [9, 12]. Sufficient flow for collapse during freeze-drying has been observed, as the viscosity of the unfrozen solute phase decreases to below  $10^4-10^7$  Pa s [19], corresponding to that of the isoviscous state at 20 to  $40^{\circ}$ C above  $T_g$  [9].

# Effects of molecular weight

The  $T_g$  of homopolymers is highly dependent on molecular weight and it increases almost linearly with the reciprocal of the molecular weight. The same applies to both  $T'_g$  and  $T'_m$  [6, 9] which, however, merge for high molecular weight carbohydrates close to 0°C [6]. The increase of the transition temperatures with molecular weight can be used in product development to improve freeze-drying behaviour and stability of the dehydrated materials. Generally addition of higher molecular weight solids increases  $T_g$ ,  $T'_g$ , and  $T'_m$ . Therefore, such materials as maltodextrins may be added in biological materials to avoid collapse and loss of quality in freeze-drying and subsequent storage.

# Conclusions

Freezing of solutions and biological materials causes freeze-concentration of solutes into an amorphous, unfrozen phase. The freeze-concentrated unfrozen solute phase solidifies into the glassy state at temperatures below the  $T'_g$  when maximum ice formation has occurred at a temperature below the onset temperature of ice melting within the maximally freeze-concentrated solute phase,  $T_m$ . Plasticization of the unfrozen phase occurs above  $T'_m$  due to ice melting, which results in significant decrease in viscosity, foaming, collapse, poor dehydration, and loss of quality. At temperatures below  $T'_m$  the freeze-concentrated solute phase can support its own weight and collapse is avoided. Therefore, the materials can be dehydrated into glassy materials which may collapse in the dehydrated state, as a result of thermal or water plasticization. State diagrams with information of conditions for equilibrium and nonequilibrium ice formation and frozen state transition temperatures can be used in selection of proper prefreezing, drying, and storage conditions. The solids composition may also be modified according to available processing and storage conditions to improve dehydration characteristics and material stability.

# References

- 1 R. J. Bellows and C. J. King, Cryobiology, 9 (1972) 559.
- 2 K. Ito, Chem. Pharm. Bull., 18 (1970) 1509.
- 3 D. Rasmussen and B. Luyet, Biodynamica, 10 (1969) 319.
- 4 L. Slade and H. Levine, Pure Appl. Chem., 60 (1988) 1841.
- 5 Y. Roos and M. Karel, Int. J. Food Sci. Technol., 26 (1991) 553.
- 6 Y. Roos and M. Karel, J. Food Sci., 56 (1991) 1676.
- 7 Y. Roos, Carbohydr. Res., 238 (1993) 39.
- 8 G. P. Johari, A. Hallbrucker and E. Mayer, Nature, 330 (1987) 552.

- 9 Y. Roos, Phase Transitions in Foods, Academic Press, San Diego, 1995.
- 10 G. P. Johari, A. Hallbrucker and E. Mayer, J. Phys. Chem., 94 (1990) 1212.
- 11 M. Sugisaki, H. Suga and S. Seki, Bull. Chem. Soc. Jpn., 41 (1968) 2591.
- 12 H. Levine and L. Slade, in J.M.V. Blanshard and J. R. Mitchell (ed.), Food Structure Its Creation and Evaluation, Butterworths, London 1988, p. 149.
- 13 F. Franks, M. H. Asquith, C. C. Hammond, H. B. Skaer and P. Echlin, J. Microsc., 110 (1977) 223.
- 14 M. Gordon and J. S. Taylor, J. Appl. Chem., 2 (1952) 493.
- 15 P. R. Couchman and F. E. Karasz, Macromol., 11 (1978) 117.
- 16 M. T. Kalichevsky, E. M. Jaroszkiewicz, S. Ablett, J. M. V. Blanshard and P. J. Lillford, Carbohydr. Polym., 18 (1992) 77.
- 17 J. L. Kokini, A. M. Cocero, H. Madeka and E. de Graaf, Trends Food Sci. Technol., 5 (1994) 281.
- 18 M. T. Kalichevsky, J. M. V. Blanshard and P. F. Tokarczuk, Int. J. Food Sci. Technol., 28 (1993) 139.